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Light-induced absorbance changes of cytochromes and carotenoids in a sulfur bacterium containing bacteriochlorophyll b

Thiococcus sp., strain Nidelven, is a photosynthetic sulfur bacterium recently isolated by Eimhjellen, Steensland and Traetteberg¹. It contains bacteriochlorophyll b as does the heterotrophic nonsulfur bacterium $Rhodopseudomonas\ viridis²$. The absorption spectra of both organisms are characterized by an extreme far-red absorption band at 1.03 μ due to bacteriochlorophyll b. The carotenoid peaks in Thiococcus¹ are located at 465, 493 and 530 nm in contrast to the peaks at 420, 452 and 482 nm in R. viridis³.

Thiococcus was grown anaerobically at 30° in an incandescent light cabinet on an inorganic medium⁴ containing Na₂CO₃ as the carbon source and Na₂S as the reductant. Absorption spectra of whole cell suspensions were recorded by a Shimadzu MPS-50L spectrophotometer. Light-induced absorbance changes were recorded by an Aminco-Chance dual wavelength spectrophotometer. Actinic light (1.01 μ) was provided by a microscope illuminator (American Optical Co., Type 653) and an interference filter (Baird-Atomic, Type B-NIR). Actinic light intensity measured with a radiometer (YSI-Kettering Model 65) was 1.5 mW·cm⁻². A 3-ml sample was pipetted directly from a culture into a 1-cm cuvette and then covered with paraffin oil for observation. The absorbance at 1.03 μ of the samples used varied between 0.1 and 0.2.

Light-induced absorbance changes were recorded at regular wavelength intervals from 380 to 600 nm. A fixed cycle of light and darkness was repeated at each measuring wavelength (e.g., 30 sec light, 90 sec dark). The light-minus-dark steady-state difference spectrum (Fig. 1) shows a peak at 408 nm and troughs at 422 nm and 556 nm, which are characteristic of the light-driven oxidation of c-type cyto-chromes in photosynthetic bacteria³. In addition, the troughs at 460, 490 and 524 nm and the peaks at 470, 505 and 543 nm indicate a light-induced red shift of the carotenoid absorption bands similar to the shifts observed previously in R. spheroides^{5,6} and Rhodomicrobium vannielii⁷. Thiococcus is the first organism containing

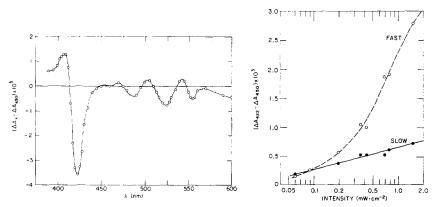


Fig. 1. Light-minus-dark difference spectrum (bandwidth = 1.5 nm).

Fig. 2. Magnitude of fast and slow phases of the transition from light to dark as a function of irradiation intensity.

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bacteriochlorophyll b to exhibit a light-induced red shift of carotenoid bands, since R. viridis does not show such a red shift³.

The kinetics of cytochrome reduction upon cessation of actinic illumination exhibited two distinct phases: a fast phase with a half time less than 2 sec and a slow phase with a half time of 20-30 sec. The fast and slow phases accounted for about 70% and 30%, respectively, of the total absorbance change at 422-425 nm after maximum intensity of actinic illumination. The difference spectrum for the slow phase (dark-minus-light) showed a peak at 421 nm and a shoulder at approx. 430 nm, whereas the spectrum of the fast phase showed evidence of only a single component at 422 nm. Light saturation curves (Fig. 2) for the two phases indicate that the cytochrome(s) involved in the slow phase reach the 50 % oxidation level at an actinic light intensity (approx. 0.2 mW·cm⁻²) less than half the intensity required for 50 % oxidation of the cytochrome(s) responsible for the fast phase (approx. 0.5 mW·cm⁻²).

In analogy to the interpretation of the cytochrome changes in the purple sulfur bacterium Chromatium⁸, the fast phase is ascribed to the reduction of cytochrome(s) in a cyclic electron transport chain, whereas the slow phase is ascribed to the reduction of cytochrome(s) by S²⁻ and related compounds via a noncyclic chain. The close similarity of the light-off kinetics and the light saturation curves for the fast and slow phases in these two sulfur bacteria support a similar interpretation for each organism.

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I K. E. EIMHJELLEN, H. STEENSLAND AND J. TRAETTEBERG, Arch. Mikrobiol., 59 (1967) 82.
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² G. Drews and P. Giesbrecht, Arch. Mikrobiol., 55 (1966) 91.

³ J. M. Olson and K. D. Nadler, Photochem. Photobiol., 4 (1965) 783.

⁴ N. PFENNIG, Zentr. Bakteriol. Parasitenk. Abt. I. Orig. Suppl., 1 (1965) 179. 5 L. SMITH AND J. RAMIREZ, J. Biol. Chem., 235 (1960) 219.

⁶ R. K. CLAYTON, Photochem. Photobiol., I (1962) 313.

⁷ S. MORITA, J. M. OLSON AND S. F. CONTI, Arch. Biochem. Biophys., 104 (1964) 346.

⁸ G. HIND AND J. M. OLSON, Ann. Rev. Plant Physiol., 19 (1968) 249.

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